

Novel Prenylated Isoflavonidal Phytoestrogen from *Trichosanthes Dioica* (Roxb)

V. K. Saxena and R.K. Dave

Natural Product Laboratory, Department of Chemistry,
Dr. H. S. Gour University, Sagar (M.P.) 470 003 India

Abstract – A novel prenylated isoflavonidal phytoestrogen glycoside; 8 – prenyl, 2'4'5' – trimethoxy isoflavone – 7 – O – β – D – glucopyranoside **1** has been isolated from ethyl acetate soluble fraction of 95% EtOH extract of leaves of *Trichosanthes Dioica* (Roxb).

Keywords – Novel Prenylated Isoflavonidal Phytoestrogen Glycoside, *Trichosanthes Dioica* (Roxb), EtOH.

INTRODUCTION

Trichosanthes Dioica Roxb [1] (Cucurbitaceae) occurring in the plains of north India and plant has been reported to be used as cardiogenic, laxative, stomachic, anti-pyretic, anti-tumor and as anti-diabetic agent (2,3). Its various parts have been phytochemically investigated by earlier workers who reported the presence of bitter principles along with amorphous saponins and essential oils. Its anti-diabetic and anti-tumor activities have already been evaluated with the significant success (4, 5).

RESULT AND DISCUSSION

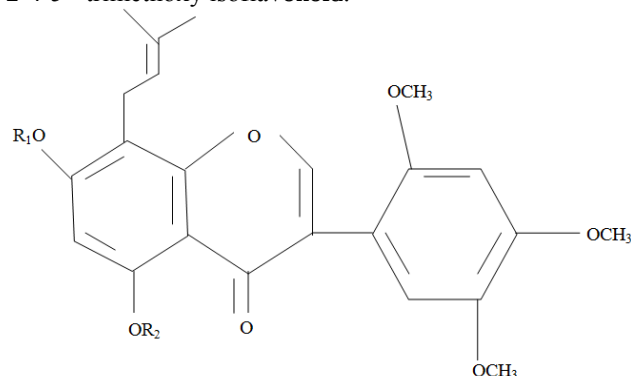
The ethyl acetate soluble extract of air dried and powdered leaves of *T. Dioica* (Roxb), on its column chromatography on Si-gel & elution with chloroform/ethylacetate furnished a light yellow coloured amorphous substance **1** m. p. 205-6°, $C_{29}H_{34}O_{12}$, M^+ 574. It gave an olive green colour with $FeCl_3$, a positive test with Na – Hg/HCl and Molish's reagent, suggesting it to be an isoflavonidal glycoside, also been supported by IR absorption at 3500, 1650, 1610, 1285, 1145, 885, 825, 766 cm^{-1} .

Acid hydrolysis of **1** yielded an aglycone **2** and sugar moieties. The sugar was identified as D-galactose. The aglycone and D-galactose were present in a equimolecular ratio (6), Aglycone **2** m.p. 247-48°, $C_{23}H_{24}O_{17}$, M^+ 412 (EIMS), gave a dark green colour with $FeCl_3$.

Preparation of its di-acetate indicated the presence of two free OH groups. A bathchromic shift of 14 & 10 nm in band II were observed on addition of $AlCl_3$ + HCl & NaOAc indicating 5, 7 di-hydroxy system in **2** (7), 1H NMR of its di acetate exhibited six proton singlet, two proton doublet & one proton triplet at δ 1.73, 3.30 (J = 7 Hz) & 5.32 respectively for prenyl unit. The downfield shift of singlet at δ 6.89 & δ 7.22 represented magnetically non – equivalent proton at C_3 & C_6 [8] whereas the sharp singlet at δ 3.50 & 2.41 were attributed for acetyl groups. These chemical shifts suggested absence of A_2B_2 system in B ring of **2**. The EIMS of **2** gave fragment ions at m/z 357 (base peak) & 356 (40 %) by the loss of M-55 & M-56, suggesting prenylation adjacent to OH group (9, 10).

Appearance of pronounced peak at m/z 381 due to M-31 indicated 2' methoxy substitution (11, 12). A peak at m/z 192 (14%) concluded trimethoxy substitution in ring B.

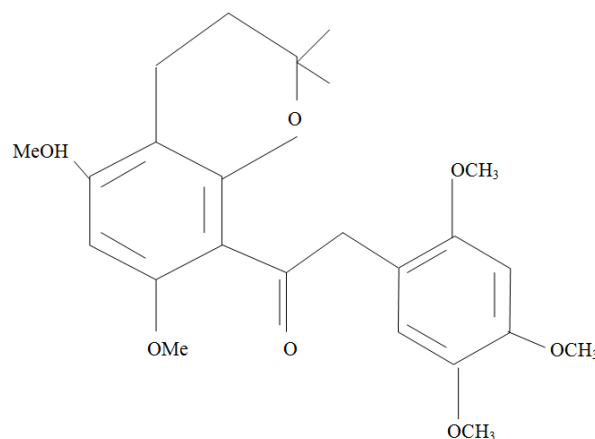
Location of prenyl unit at C-8 position in ring A was established by the significant bathchromic shift 55 nm on addition of $AlCl_3$ (13). On mild alkaline hydrolysis followed by cyclization of dimethyl ether of **3** yielded **2**; 2, dimethyl chromant identified by PMR (14) also conforming prenylation of C_8 . Further $KMnO_4$ oxidation of dimethyl ether of **2** formed 2, 4, 5 tri-methoxy benzoic acid thereby conforming methoxyl position in ring B, thus the aglycone **2** was identified as 5, 7 dihydroxy 8 prenyl 2'4'5' trimethoxy isoflavonoid.



1, R_1 = D-galactose

2, R_1R_2 = H

3, R_1R_2 = Me



Periodic oxidation and enzymatic hydrolysis (15) revealed that the glycoside **1** consists of one mole of the aglycone and one mole of D-galactose and linkage was β . The glycoside did not show the bathchromic shift on addition of NaOMe suggesting the glycosylation of the OH group in ring A. The permethylation (16) confirmed

that C₁-OH of sugar was attached with C₇ - OH of aglycone, thereby confirming that the 1 as 8-prenyl - 2', 4', 5' - trimethoxy isoflavone - 7 - O - β - D - galactopyranoside.

The isolated compound 1 was found to be estrogenically active when administrated (17) on albinorats because of the fact that it led to the increase in the weight of uterine.

EXPERIMENTAL

PLANT MATERIAL: T. Dioica (Roxb) was procured from M/s. United Chemical and Allied Products, Calcutta. The plant was authenticated by the Department of Botany, courtesy of Dr. Harisingh Gour University, Sagar, M.P. A herbarium specimen (No. IV - XVII) has been deposited in the Department of Chemistry of the University.

ISOLATION: Air-dried powdered leaves (4.0 kg) of T. Dioica were extracted exhaustively with hot 95% ethanol. The extract was concentrated under vacuum & segregated into petroleum ether, C₆H₆, EtOAc, (Me)₂CO soluble fractions. The ethyl-acetate soluble fraction showed the presence of two spots on TLC. These were separated by column chromatography on Si gel (60-120 mesh) column and eluted with chloroform / ethyl-acetate in various proportions.

COMPOUND 1: Eluate with CHCl₃ / MeOH (5:6), light yellow amorphous compound m.p. 205-206, mf. C₂₉H₃₄O₁₂ (found C 60.59, H 5.91, Calcd. C 60.62, H 5.92). M⁺ 574 (EIMS), intense green colour with FeCl₃ pink colour with Na-Hg / HCl & positive molish's test. UV λ_{max}^{MeOH} 262, 291 (sh), 300 (sh), $\lambda_{max}^{AlCl_3}$ 275, 310, $\lambda_{max}^{AlCl_3/HCl}$ 276, 309, ; IR ν_{max}^{KBr} 3500, 2930, 2820, 1650, 1625, 1557, 1570, 1385, 1368, 1285, 1215, 1145, 885, 825, 766, 710 . & 690 cm⁻¹; fabms m/z 574 (1), 412 (23), 384 (42), 381 (56), 357 (31), 356 (74), 192 (93), 166 (100).

ACETYLATION OF COMPOUND 1: Compd 1 heated with fused sodium acetate and acetic anhydride at 130° in oil bath for 4 hrs & workup as usual & acetyl derivative crystallized with methanol white needles, m.p. 185-86° analysed for C₃₉H₄₄O₁₇ (found C 59.68, H 5.59, Calcd. C 59.69, H 5.61). IR ν_{max}^{KBr} 1720, 1645 cm⁻¹. PMR (CDCl₃ 90 MHz, TMS ppm) 8.02 (s, H, C₂), 7.23 (s, H, C₆), 6.82 (s, H, C₃), 5.32 (t, H, C_{2''}), 3.92 (s, 3H, OCH₃), 3.82 (s, 3H, OCH₃), 3.79 (s, 3H, OCH₃) 3.30 (d, J = 6 Hz, H, C_{1''}), 1.72 (s, 6H, C_{4''}, C_{5''}), 5.16 (d, J = 7 Hz, H, C_{1''}), 4.3-4.8 (m, 6H of galactose), 2.52 (s, 3H, OAc C_{6'''}), 2.14 (s, 3H, OAc, C_{4'''}), 2.05 (s, 3H, OAc, C_{2'''}).

ACID HYDROLYSIS OF 1: 1 was refluxed with 7 % alc. H₂SO₄ for 7 hrs. The alcohol was removed and a cream coloured substance 2 was separated. The concentrated hydrosate after neutralization, was examined on PC (BAW 4:1:5) & found to contain galactose. The quantitative estimation of sugar showed that the aglycone & galactose are present in a equimolecular ratio.

COMPOUND 2: Cream coloured substance recrystallised with MeOH : acetone, m.p. 247-48°, analysed for C₂₃H₂₄O₇ (found C 66.98, H 5.81, Calcd. C

66.99, H 5.82), M⁺ 412 (EIMS), gave dark green colour with FeCl₃, Pink colour with Na-Hg / HCl. UV λ_{max}^{MeOH} 254, 290(sh), 307(sh), $\lambda_{max}^{AlCl_3}$ 271, 309, $\lambda_{max}^{AlCl_3+HCl}$ 273, 306 ; IR ν_{max}^{KBr} 3510, 2915, 2830, 1640, 1560, 1500, 1380, 1367, 1240, 1190, 805cm⁻¹. Fams m/z (r.int) 412 (24), 384 (40), 381 (57), 357 (29), 356 (70), 192 (91), & 166 (100). ¹³C NMR (DMSO - d₆, 22.6 MHz), 153.6 (C-2), 120.1(C-3), 178.6 (C-4), 158.4 (C-5), 98.4 (C-6), 161.1 (C-7), 154.3 (C-9), 104.8 (C-10), 108.8 (C-1'), 155.6 (C-2'), 102.4 (C-3'), 159.1 (C-4'), 105.6 (C-5'), 132.1 (C-6'), 22.4 (C-1''), 123.3(C-2''), 129.6 (C-3''), 25.1 (C-4''), 17.3 (C-5'') 60.2 (20 CH₃), 55.1 (4' OCH₃), 55.9 (5' OCH₃).

ACETYLATION OF 2: The acetyl derivative of 2 was prepared by sodium acetate and acetic anhydride and obtained as silk needled, m.p. 226-27° C₂₇H₂₈O₉ (found C 65.31, H 5.62, Calcd. C 65.32, H 5.64), IR ν_{max}^{KBr} 1750, 1645 cm⁻¹; PMR (90 MHz, CDCl₃ ppm), 8.02 (s, H, C₂), 7.23 (s, H, C₆), 6.87 (s, H, C₃), 5.32 (t, H, C_{2''}), 3.94 (s, 3H, OCH₃), 6.87 (s, 3H, C₃), 5.32 (t, H, C_{2''}), 3.94 (s, 3H, OCH₃) 3.79 (s, 3H, OCH₃), 3.30 (d, J = 7 Hz, 2H, C_{1''}), 2.50 (s, 3 H, OAc, C₅), 2.41 (s, 3H, OAc, C₇), 1.73 (s, 6H, C_{4''}, C_{5''}).

METHYLATION OF 2 TO 3: Compound 2 (400mg) + DMSO₄ + anhydrous K₂CO₃ + dry acetone were refluxed for 8 hours. The inorganic salts were filtered off, acetone was evaporated to a syrupy mass & it was charged over small silica gel column & eluted with ether. The removal of ether & on crystallization it yielded white needles (260 mg) m.p. 216-17° analyzed for C₂₅H₂₈O₇ (found C 66.17, H 6.34, Calcd. C 68.18, H 6.36).

ALKALINE HYDROLYSIS AND CYCLIZATION: The di-methyl ether 3 (200 mg) in 20% alco. NaOH (10 ml) refluxed for 20 minutes. The reaction mixture was acidified after cooling and water was added & extracted with ether. The ether soluble part on evaporation yielded a residue which was mixed with HCOOH (5 ml) & kept for an hour, followed by addition of H₂O and was extracted with ether. Removal of ether from ether soluble part yielded a gummy substance, analyzed for C₂₄H₃₀O₇ and was identified as 2, 2 di-methyl chroman.

KMnO₄ OXIDATION OF 3: Potassium permanganate was added in small quantities to 3 in acetone. The mixture was refluxed and more KMnO₄ was added until slight excess was present. The mixture was refluxed for further 15 minutes. The precipitated manganese dioxide was filtered off and washed with H₂O. The combined filtrate and washing were acidified with H₂SO₄, saturated (NH₄)₂SO₄ & extracted with ether. Removal of ether and crystallization of product yielded 2, 4, 5 tri-methoxy benzoic acid.

PERIODATE OXIDATION: Compound 1 was dissolved in methanol (40 ml) and kept with sodium meta-periodate (15 ml) in a conical flask for 2 days. The liberated formic acid and consumed periodate was estimated by Jone's method.

PERMETHYLATION AND ACID HYDROLYSIS: Compound 1 was treated with CH₃I & Ag₂O in dimethyl formamide at room temperature. After 2 days, the reaction

mixture was filtered and the residue was washed with dimethyl formamide. The filtrate was concentrated in a vacuum and hydrolysed by alc. H_2SO_4 . After usual workup methylated sugar was identified on Pc as 2,3,4,6 tetra methyl galactose.

ACKNOWLEDGEMENT

We are thankful to the director, CDRI, Lucknow, for recording various spectra, and to Head of Chemistry, Deptt. of this University for providing laboratory facilities.

REFERENCES

- [1] "Wealth of India "A Dictionary of Indian Raw Material and Industrial product, C.S.I.R. Pub. New Delhi, Vol. X, p. 473 (1976).
- [2] Chopra, R.N. Nayer, S.L. Chopra, I.C. , Glossary of Indian Medicinal Plants, C.S.I.R. Pub., New Delhi, p. 248 (1980).
- [3] Kritikar , K.K. and Basu , B.D. , Indian Medicinal Plants , Lalit Mohan Basu Pub., Allahabad, Vol. I.,p. 581 (1918).
- [4] North, B. and Roy, A.C., Patna University J. , Vol. I, No. 2. p. 56-60 (1945).
- [5] Sharma, Govind and Pant, M.C. Current Science, Vol. 57, No. 19 , p. 1085(1988)
- [6] Mishra, S.P. & Mohan Rao V.K., J. Sci. Ind. Res. Sect. C 19, 170 (1960).
- [7] Mabry, T.J., Markham, K.R. & Thomas , M.B., "The Systematic Identification of Flavonoids ", p.170-71, Springer Verlag , New York.
- [8] Sharma , A., Chibber and Chawla , H. M. , Ind. J. Chem. , 18 B , 472 (1979).
- [9] Ritchie, E., Taylor , W. C. and Williams , D.H. , Tet. Lett. , 1437 (1972).
- [10] Subbarao A. V., Rathi , S.S. and Venkataraman , K. , Ind. J. Chem. , 10, 989 (1964).
- [11] Reed , R. I. and Wilson , J. M. , J. Chem. Soc. 5949 (1963).
- [12] Campbell, R.V.M., Harper, S.H. and Kemp, A.D., J. Chem.Soc. (C) , 1787 (1969).
- [13] Tokuoka, Y. Daigo , K. and Takemoto , T. " Yakugaku Zasshi", 95 , 825 (1975).
- [14] Falshaw, C.P., harmer, R.A., Ollis, W.D., Wheeler, E., Lalitha, V. R. and Subba Rao, N.V., J. Chem. Soc.(C), 374 (1969).
- [15] Hirst, E.L. and Jones ,J.K.N., J.Chem Soc., 1959 (1949).
- [16] Khun, R., Trichamann, H and Low, I . "Angew Chem.", 32, 67 (1955).
- [17] Bickoff, E.M., Livingston and Booth, A.N., J.Pharm. Sci., 1411 (1964).